

**Identification of the Ecological Niches and Development of  
Intervention Strategies to Reduce Pathogenic Foodborne  
Pathogens in Poultry**

**CRIS Project 3091-32000-035-00D**

**Michael Kogut, Lead Scientist**

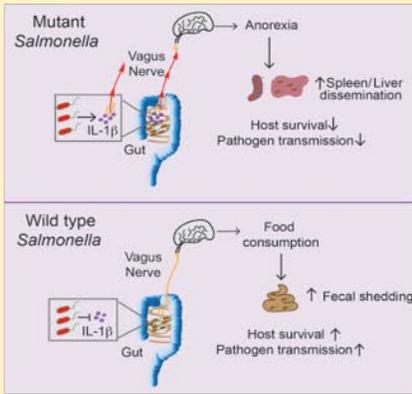
**Kenneth Genovese, Christina Swaggerty, Haiqi He,**

**Toni Poole, Michael Hume**

## **2nd International Symposium on Alternatives to Antibiotics was held at the World Organization for Animal Health (OIE)**

- **Vaccines**
- **Bacterial-derived products**
- **Phytochemicals**
- **Immune-related products**
- **Innovative drugs, chemicals, and enzymes**

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**Immune response/persistence:**  
**Disease resistance vs disease tolerance**



5

**Gut health:**  
**Microbial metabolites**  
**Dietary additives**

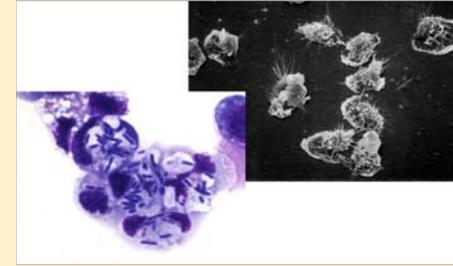
**Proteomics & metabolomics:**  
**Immunometabolic reprogramming**  
**Epigenetics**

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**Innate Immunity-  
 Based Vaccines:**  
**TCA mutants**  
**Salmonella protein array**



**Modulation of  
 innate immunity:**  
**Trained immunity**

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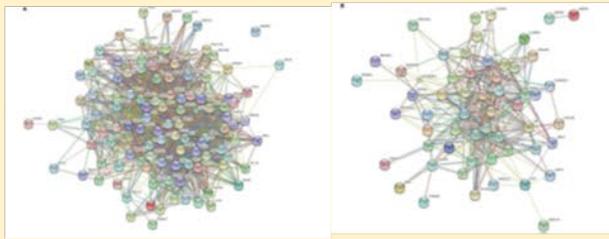
**Molecular Interplay  
 Between the Avian Host  
 and Salmonella at the  
 Gut Interface**

**Going Forward** →

**IMPACT**

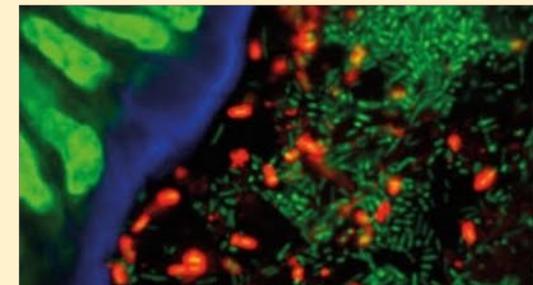
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**Immunogenetics:**  
**Protein:protein interactions**



**Microbiota & mucosal immunity:**  
**Bacterial taxa & cytokine gene expression**  
**Microbial metabolite modulation**

4



### Infection via faecal-oral route

- Attachment and invasion of intestinal epithelium
- Activation of innate immune response through action of bacterial effector proteins and host recognition (TLR 4, 5 & 21)
- Pro-inflammatory CXC chemokine response leads to influx of heterophils
- Damage to intestinal epithelium but activation of response
- Host-adapted serovars (*S. Gallinarum* & *S. Pullorum*) invade via 'stealth' infection-little or no activation of innate response
- Absence of flagella in adapted serovars avoids TLR5 recognition & targeting of lymphoid tissue (caecal tonsils)

Initial Infection

### Systemic response

- Translocation to spleen and liver
- Establishment of intracellular infection in macrophages
- Macrophage activation and antimicrobial activity related to genetic resistance encoded by *SAL1* locus
- Initiation of cellular response-key role in IFN- $\gamma$  by  $\gamma\delta$  T cells
- Immunomodulation through up-regulation of  $T_H2$  response by avian adapted serovars
- Development of IgM and IgY antibody response

### Gastrointestinal Response

- Initial inflammatory response regulated by regulatory T cells
- Role of  $T_H17$  response in maintaining gut integrity?
- Mucins and gallinacins limit infection
- Secretory IgA response
- *Salmonella* persistence within lower intestinal tract (caeca)

Establishment of Infection

### Death

- Uncontrolled replication in macrophages of susceptible birds leading to bacteraemia

### Clearance

- Systemic clearance at 2-4 weeks post infection by both cellular and antibody responses
- Intestinal clearance 3-12 weeks dependent on  $T_H1$  response
- Depletion of antibody through bursectomy has no effect on clearance

### Persistence

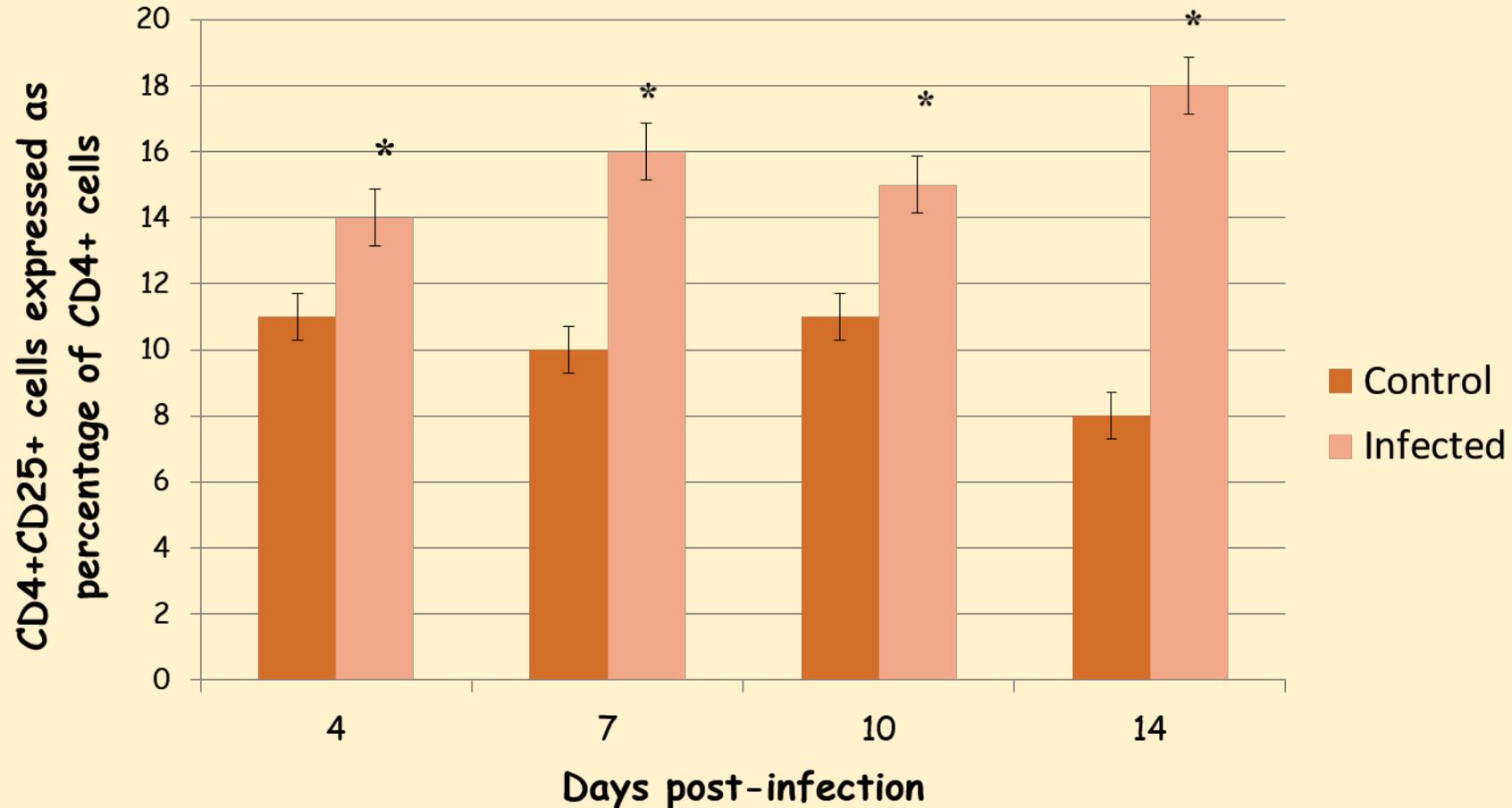
- Intracellular persistence within macrophages by adapted serovars
- Recrudescence of infection in hens following immunosuppression due to a drop in CD4<sup>+</sup> cell numbers at sexual maturity

Outcome

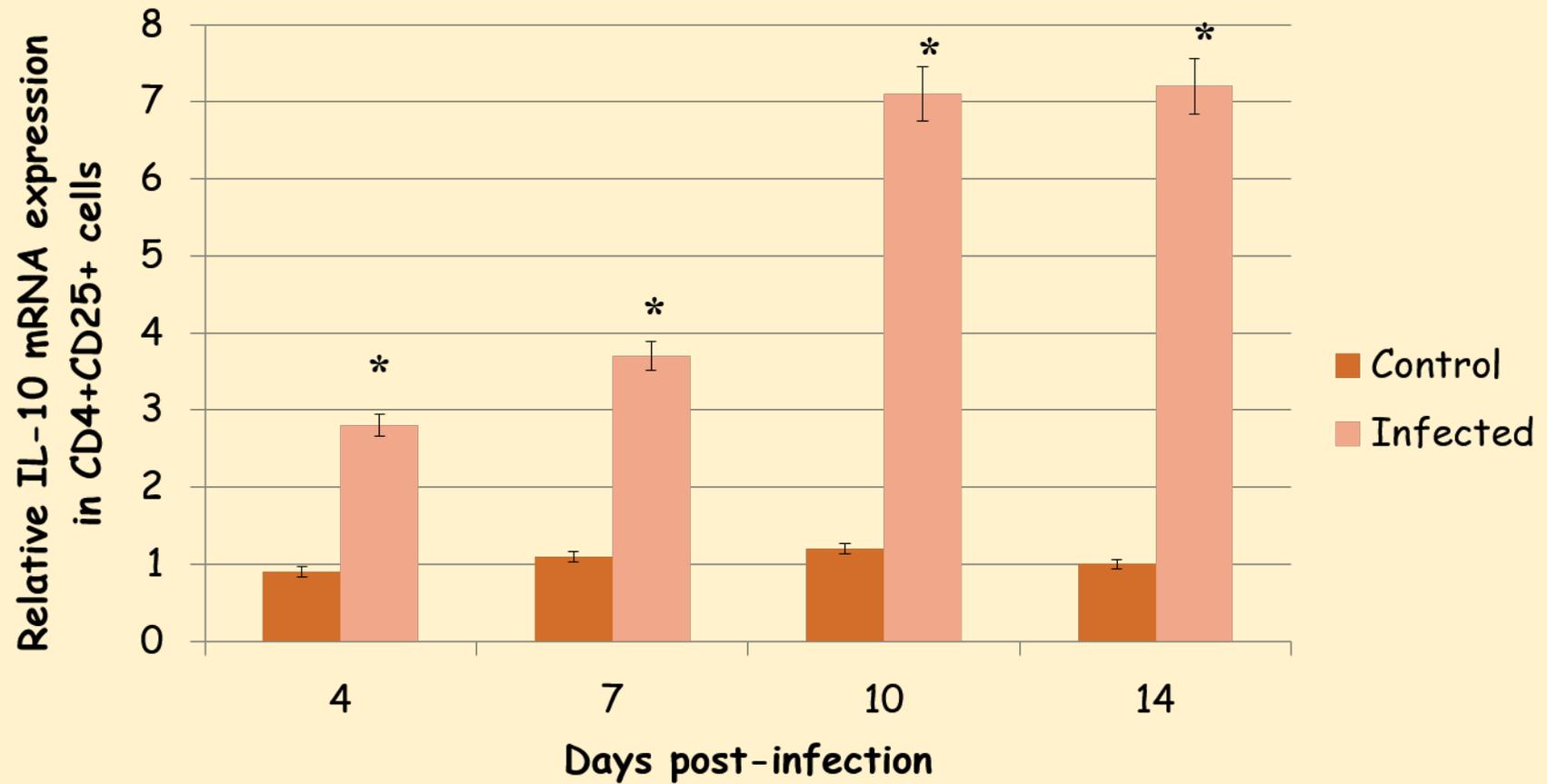
# Persistence of *Salmonella* in Intestine of Broilers

- A. What are the mechanisms that underlie persistence of *Salmonella* in the chicken gut?**
- Host immunity
  - Local tissue metabolism
  - Bacterial effectors
  - Microbiota metabolism
- B. What regulates the GI response to prevent excessive intestinal damage?**
- Host immunity
  - Local tissue metabolism
  - Bacterial effectors

# Percentage of CD4+CD25+ cells in cecal tonsils following *Salmonella* Enteritidis infection



## IL-10mRNA expression in Tregs following infection with *Salmonella* Enteritidis



# Phosphorylation mediated signal transduction

- Phosphorylation = protein PTM
  - virtually every cellular process regulated by phosphorylation
- Phosphorylation events are catalyzed by kinases
- Estimated that a third of proteins are modified through phosphorylation.

**Understanding cellular kinase activity (*the kinome*) can offer insight into complex biology, identify biomarkers associated with valuable phenotypes and provide therapeutic targets**

# Kinomics

- 530 kinases in chicken genome
- 100,000 unique phosphorylation sites in chicken proteome
- Diversity of kinases and targets in different species

## FUNCTIONAL PHENOTYPE

Signaling pathways activate/deactivated during treatment, disease, insult

# Immunometabolic Kinome Array

- 1000 peptides
- Species-specific
  - **Chicken**
  - Turkey
  - Bovine
  - Porcine
- Process-specific
  - Innate & acquired immune responses
  - Protein, CHO, FA metabolism
  - Hormone & stress responses

# Pathometabolism

- Metabolic switches targeted by pathogens
    - ATP-activated protein kinase (AMPK)
    - Hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ )
    - Mechanistic target of rapamycin (mTOR)
- Metabolic checkpoints  
"core" cell signaling pathways
- Re-program host cell metabolism – nutritional requirements
  - Epigenetic modifications: long term alterations in gene expression

# Immuno-pathometabolic signatures in the cecum of chickens infected with *Salmonella enterica* serovar Enteritidis

Characteristic	Early (4-72 hours)	Late (4-14 days)
Immune cells	Heterophils, macrophages	Tregs
Transcription factor activation	NF-κB, JAK2, JAK3, STAT4	NFAT, Smad1-3, STAT1, STAT6
Cytokine mRNA expression	IL-1β, IL-6 IFN-γ	IL-10, TGF-β4
Metabolism	Anabolic	Catabolic
Characteristic metabolic processes	Glycolysis, protein synthesis	Oxidative phosphorylation
Energy balance	AMP:ATP	AMP:ATP
Regulatory enzyme activity	mTORC1 phosphorylation	AMPKa phosphorylation
Tissue phenotype	Pro-inflammatory	Anti-inflammatory
	<b>Disease resistance</b>	<b>Disease tolerance</b>

# Salmonella infection in cecum= *Skeletal muscle*

- **Initial response (24-96 h) is anabolic:**
  - glycolysis & FA oxidation
  - protein synthesis
  
- **Between 4-21 days, metabolism turns catabolic:**
  - AMPK $\alpha$  phosphorylation = FA synthesis, lipogenesis
  - Insulin = increase in glucose transport

# Host Defense Mechanisms

## Disease resistance

Mediated by immune response

Reduce invasion

Reduce/eliminate pathogens

- mechanisms to control pathogen invasion & replication
- reducing pathogen load and hence pathogenicity, but with immunopathology

## Disease tolerance

Limit damage

No affect on growth/numbers

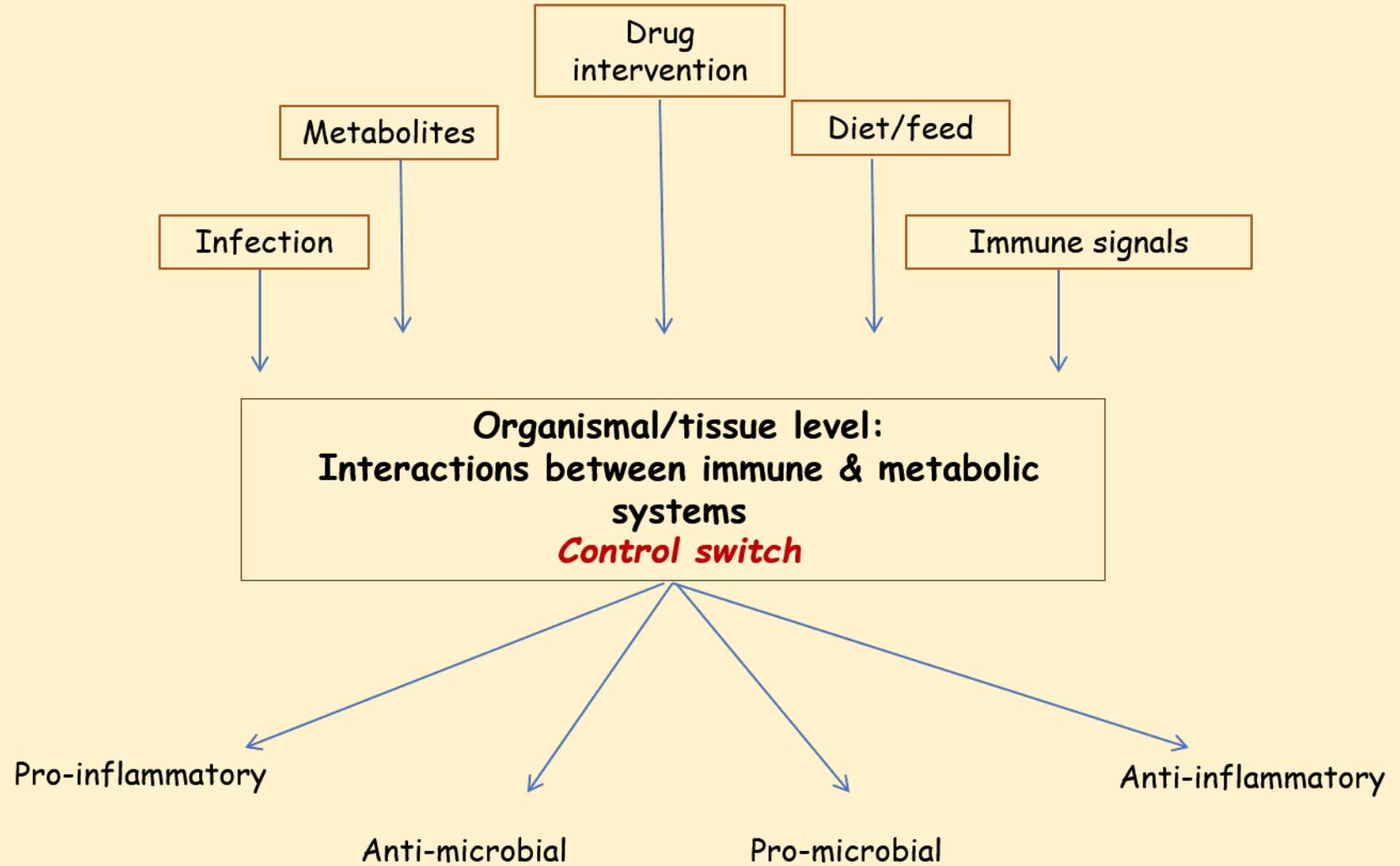
Different mechanisms

- mechanisms to control the damage of infection & resulting IR
- reduces disease severity w/o interfering with pathogen load

# IMPACT

- Pathometabolism - Disease tolerance
- Emerging functions of the immune system in regulating tissue physiology
- Identified host regulatory molecules and pathways modulated during *Salmonella* persistence
  - inhibitory pathways hardwired into physiological systems (immunity, metabolism)
  - crucial for maintaining and modulating the duration and amplitude of physiological responses in peripheral tissues in order to minimize collateral tissue damage

**Input**



**Output**

**Targeting Metabolic Pathways to Tune Immunity:  
harness immunometabolism for novel drug discovery**

# Future OSQR Plans

- **Role of *Salmonella***

- *Icd, Acn* (TCA cycle) ]- Inhibit NLRP3 Inflammation activation
- *SlrP*
- *SarA* – *Salmonella* anti-inflammatory response activator = STAT3-dependent IL10
- Consistent between paratyphoid serovars

- **Role of microbiome**

- Gut phenotype – microbiome function
- Diversity
- Metabolite production = metabolic reprogramming

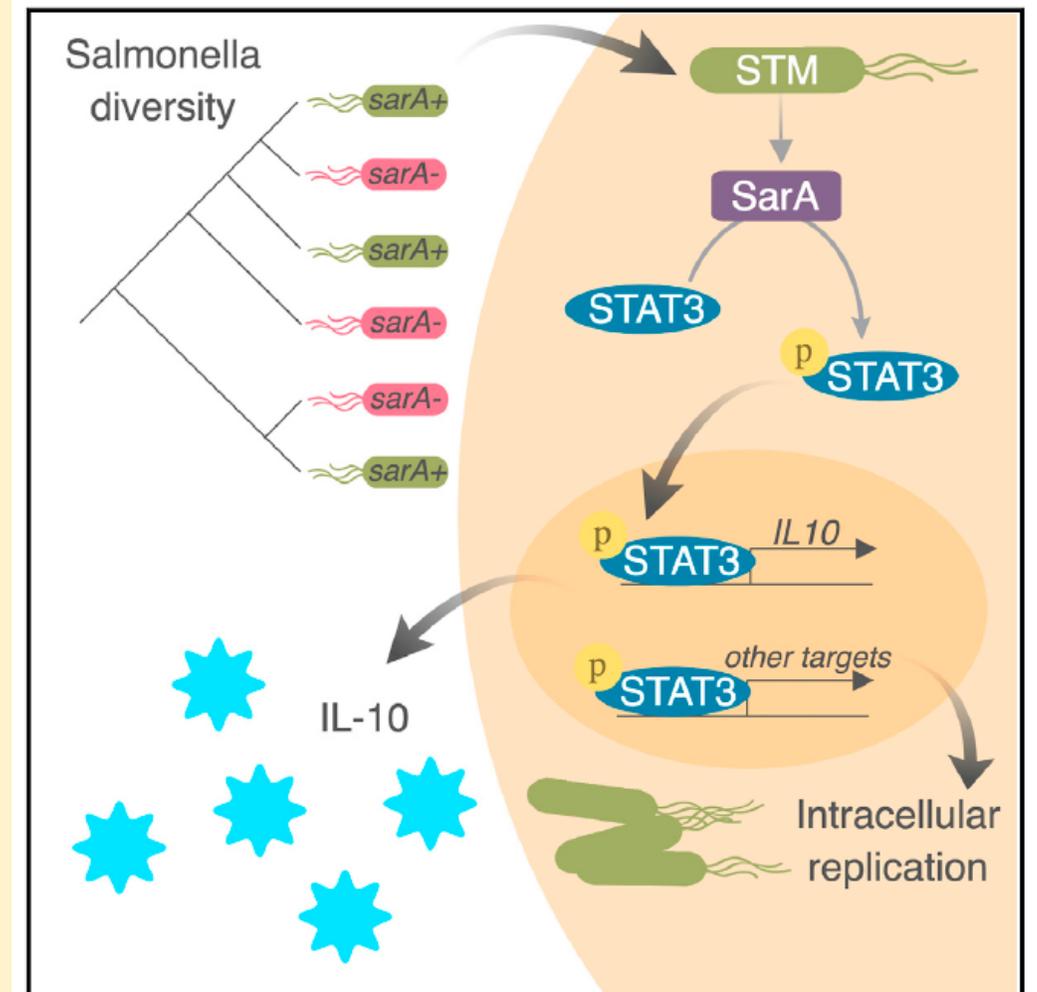
- **Role of the host**

- Gut-brain axes: vagus nerve and IL1B-dependent genes
- Epigenetics: metabolic reprogramming

# Salmonella Activation of STAT3 Signaling by SarA Effector Promotes Intracellular Replication and Production of IL-10

## Highlights

- SarA (*Salmonella* anti-inflammatory response activator) effector induces IL-10
- SarA activates an anti-inflammatory STAT3 transcriptional program
- SarA contributes to *Salmonella* intracellular replication and virulence in mice
- Variable presence in strains facilitated SarA identification, suggests adaptation



## **Vaccination and early protection against non-host-specific *Salmonella* serotypes in poultry by exploiting innate immunity**

- **Recognition of PAMPs by specific PRRs triggers innate immunity, which induces a variety of gene expression via distinct intracellular signaling pathways**
- **Pattern recognition receptors:**
  - TLRs: the plasma membrane or endosomes
  - NLRs: within the cytosol of most cell types in intestinal mucosa.

## Oxidative metabolism enables *Salmonella* evasion of the NLRP3 inflammasome

Meghan A. Wynosky-Dolfi,<sup>1</sup> Annelise G. Snyder,<sup>1</sup> Naomi H. Philip,<sup>1,2</sup> Patrick J. Doonan,<sup>1</sup> Maya C. Poffenberger,<sup>6,7</sup> Daina Avizonis,<sup>7,8</sup> Erin E. Zwack,<sup>1,3</sup> Amber M. Riblett,<sup>3</sup> Baofeng Hu,<sup>1</sup> Till Strowig,<sup>4</sup> Richard A. Flavell,<sup>4,5</sup> Russell G. Jones,<sup>6,7</sup> Bruce D. Freedman,<sup>1,2</sup> and Igor E. Brodsky<sup>1,2,3</sup>

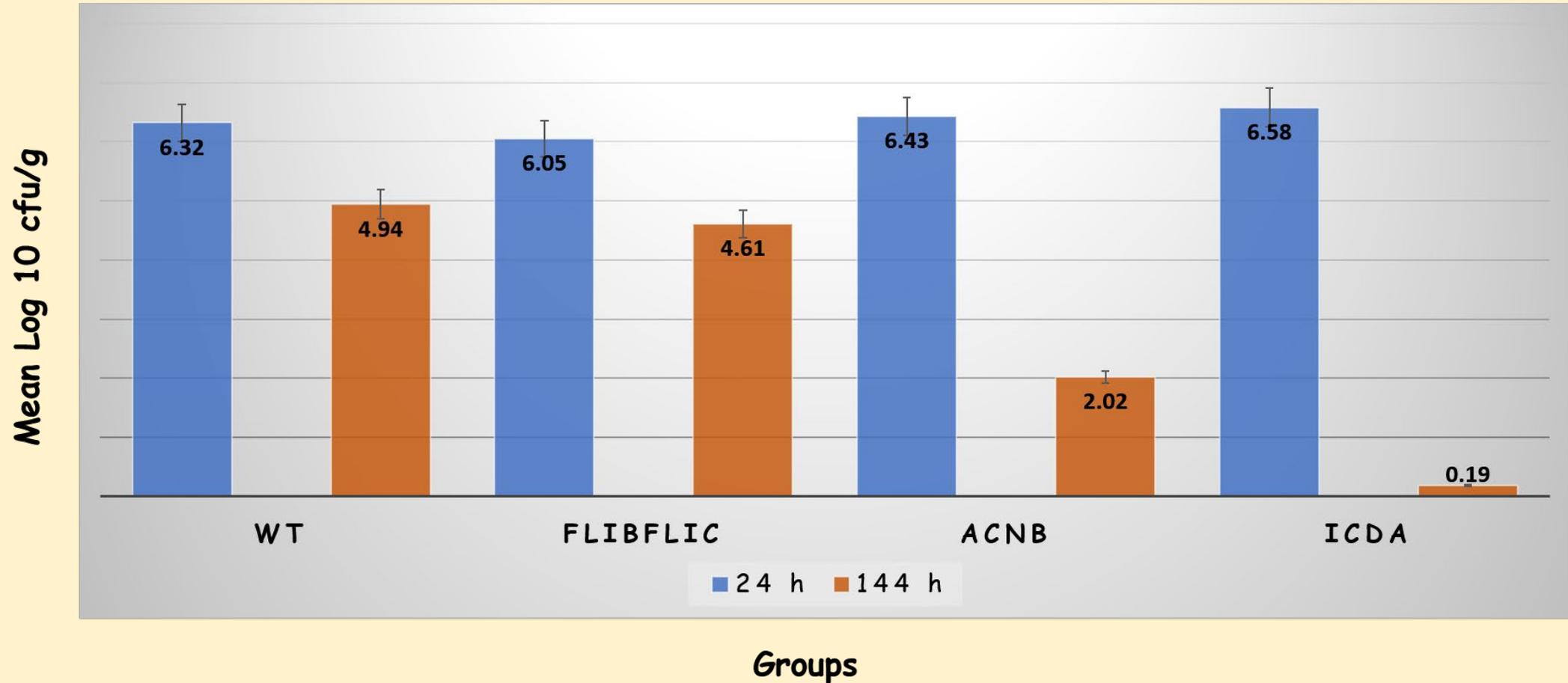
<sup>1</sup>Department of Pathobiology, School of Veterinary Medicine; and <sup>2</sup>Immunology Graduate Group and <sup>3</sup>Cell and Molecular Biology Graduate Group, University of Pennsylvania, Kennett Square, PA 19104

<sup>4</sup>Department of Immunobiology and <sup>5</sup>Howard Hughes Medical Institute, Yale University School of Medicine, New Haven, CT 06510

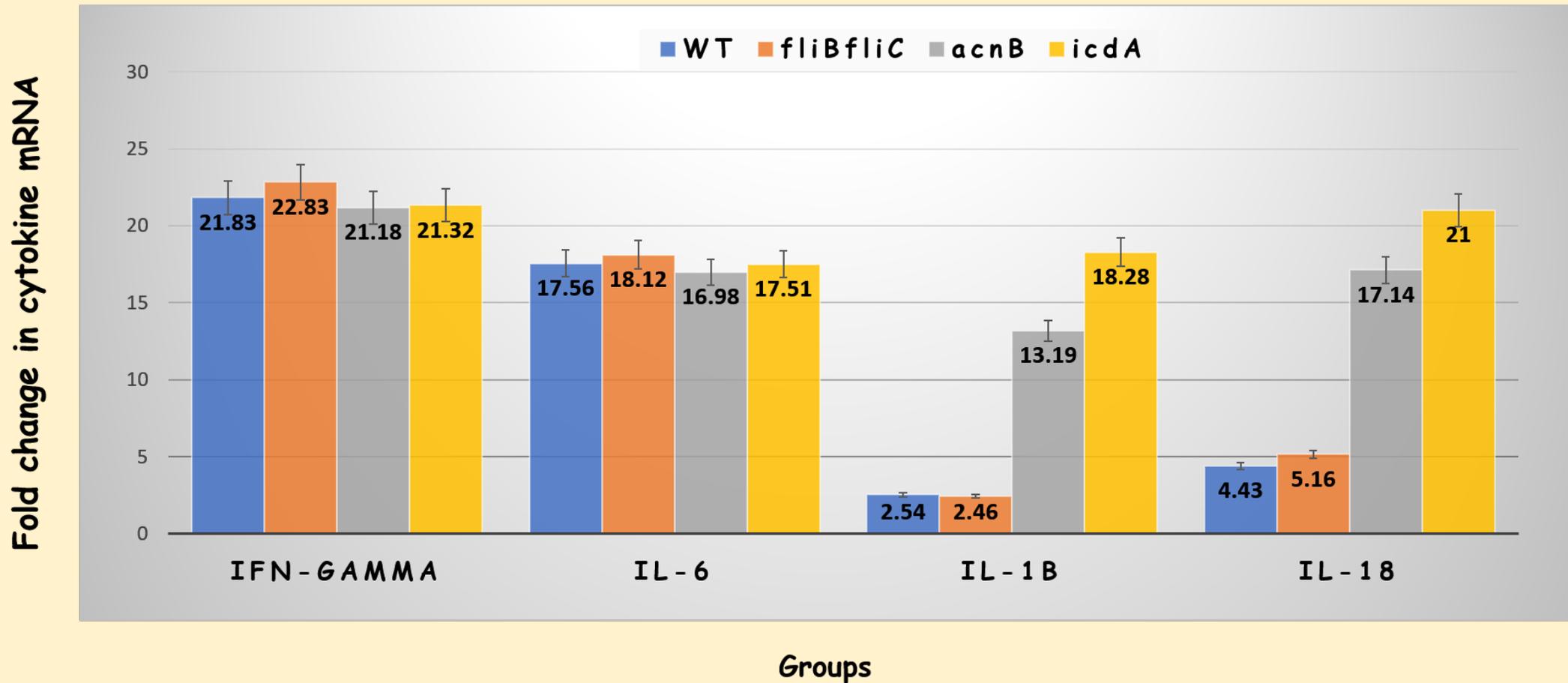
<sup>6</sup>Department of Physiology, <sup>7</sup>Goodman Cancer Research Centre, and <sup>8</sup>Metabolomics Core Facility, McGill University, Montreal, Quebec H3A 0G4, Canada

identify bacterial factors that limit NLRP3 inflammasome activation. Surprisingly, absence of the *Salmonella* TCA enzyme aconitase induced rapid NLRP3 inflammasome activation. This inflammasome activation correlated with elevated levels of bacterial citrate, and required mitochondrial reactive oxygen species and bacterial citrate synthase. Importantly, *Salmonella* lacking aconitase displayed NLRP3- and caspase-1/11-dependent attenuation of virulence, and induced elevated serum IL-18 in wild-type mice. Together, our data link *Salmonella* genes controlling oxidative metabolism to inflammasome activation and suggest that NLRP3 inflammasome evasion promotes systemic *Salmonella* virulence.

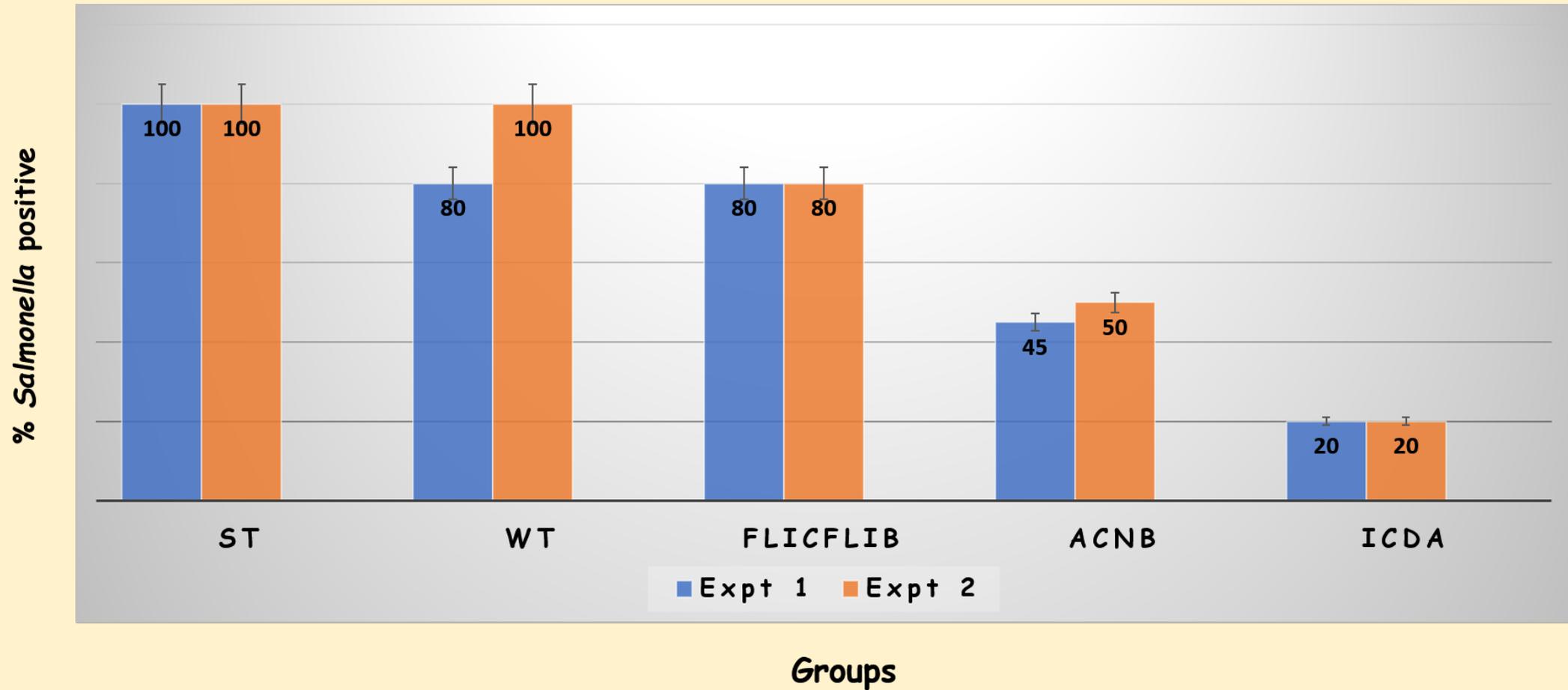
# Cecal bacterial CFU post-infection (log 10) (24 and 144 h p.i.)



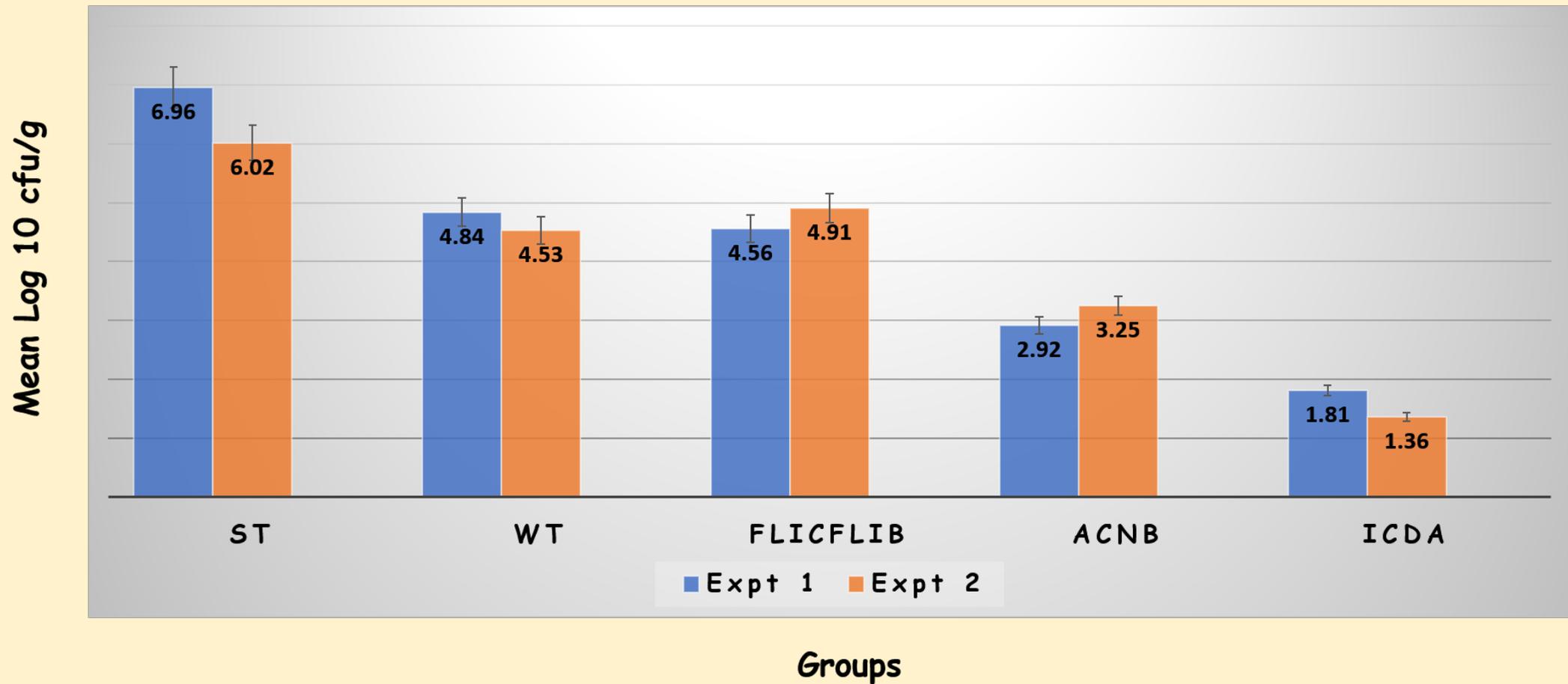
# Fold Change in Cytokine mRNA Expression in Cecal Tissue (24 h p.i.)



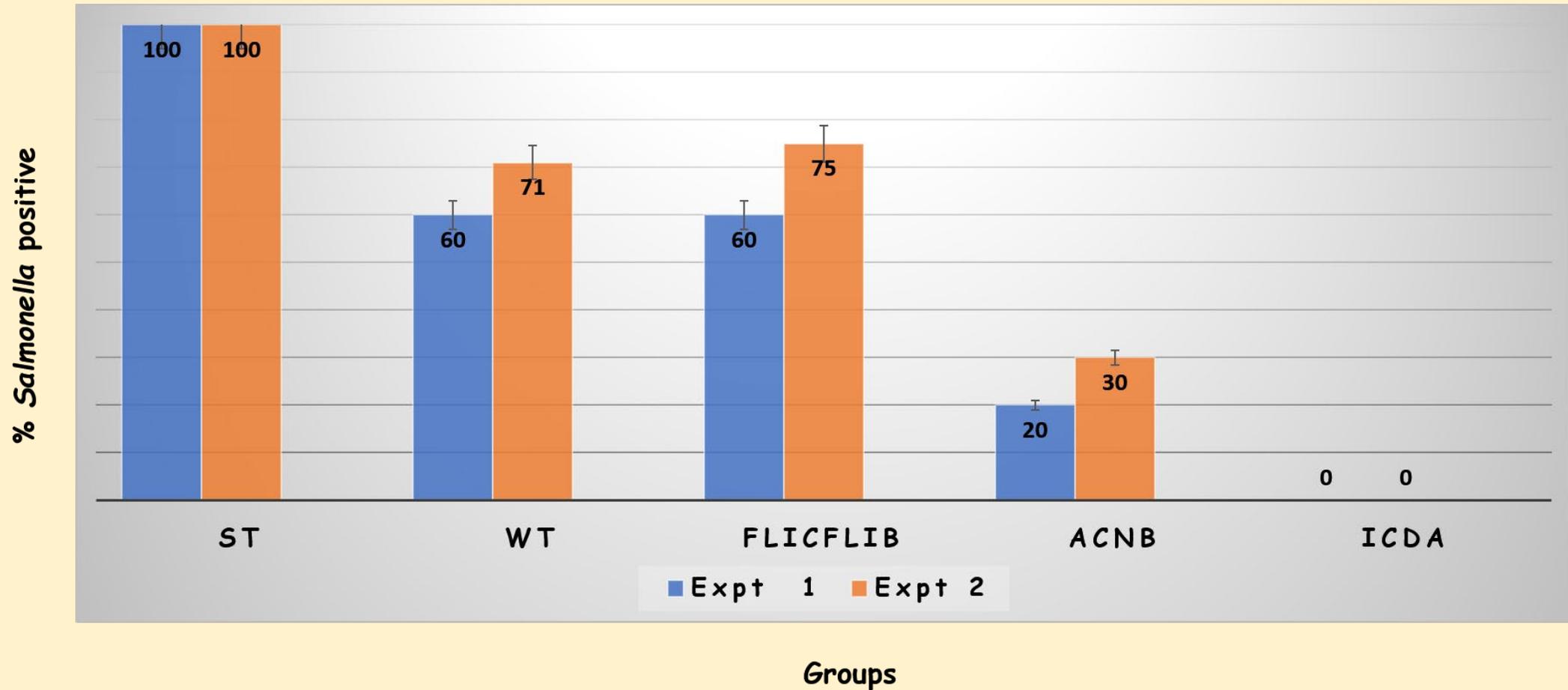
**Percent positive cecal colonization  
(7 days post-challenge)  
(challenge 3 weeks post immunization)**



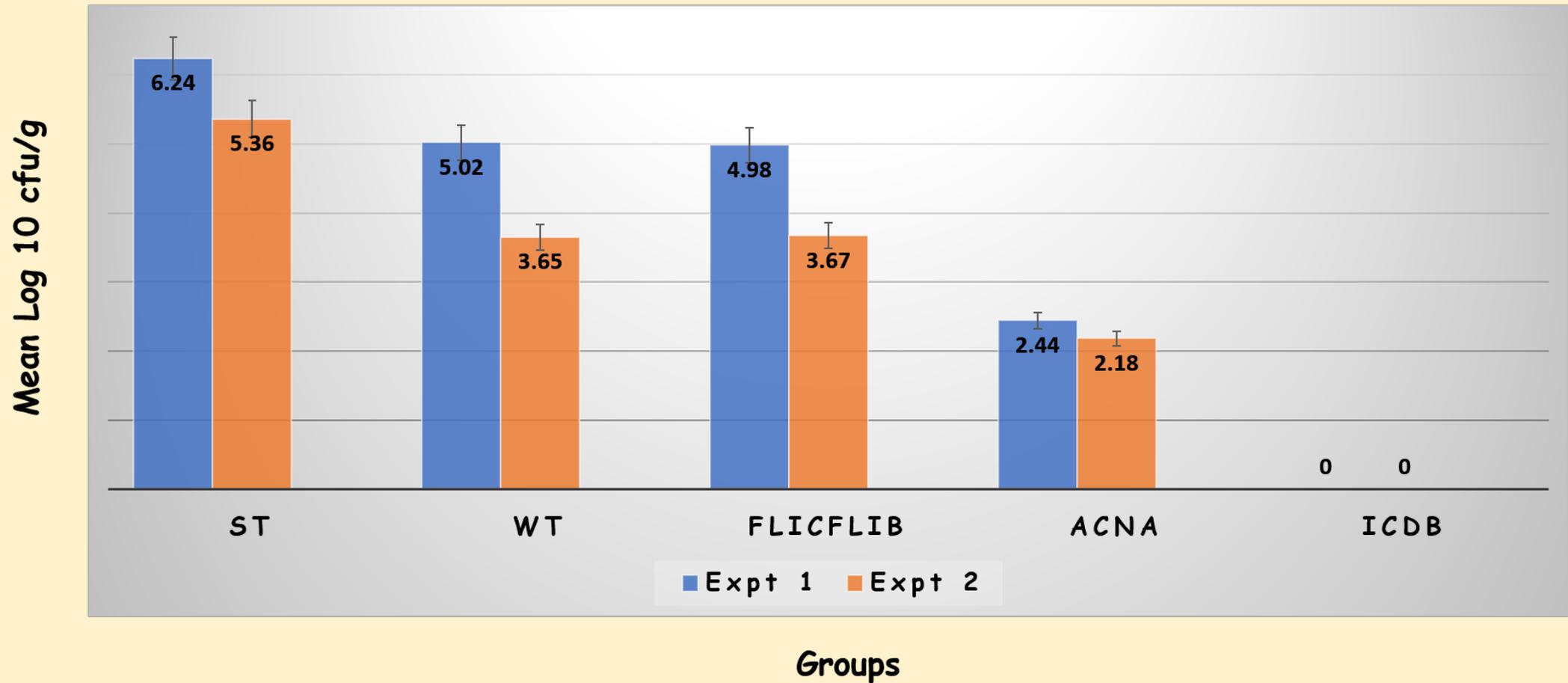
**Bacterial cecal (cfu/g) colonization  
(7 days post-challenge)  
(challenge 3 weeks post immunization)**



**Immunization trials:  
Organ invasion (2 days post-challenge with chicken ST)  
(challenge 3 weeks post immunization)**



**Immunization trials:  
Liver/spleen cfu/g (2 days post-challenge with chicken ST)  
(challenge 3 weeks post immunization)**



# Impact (2)

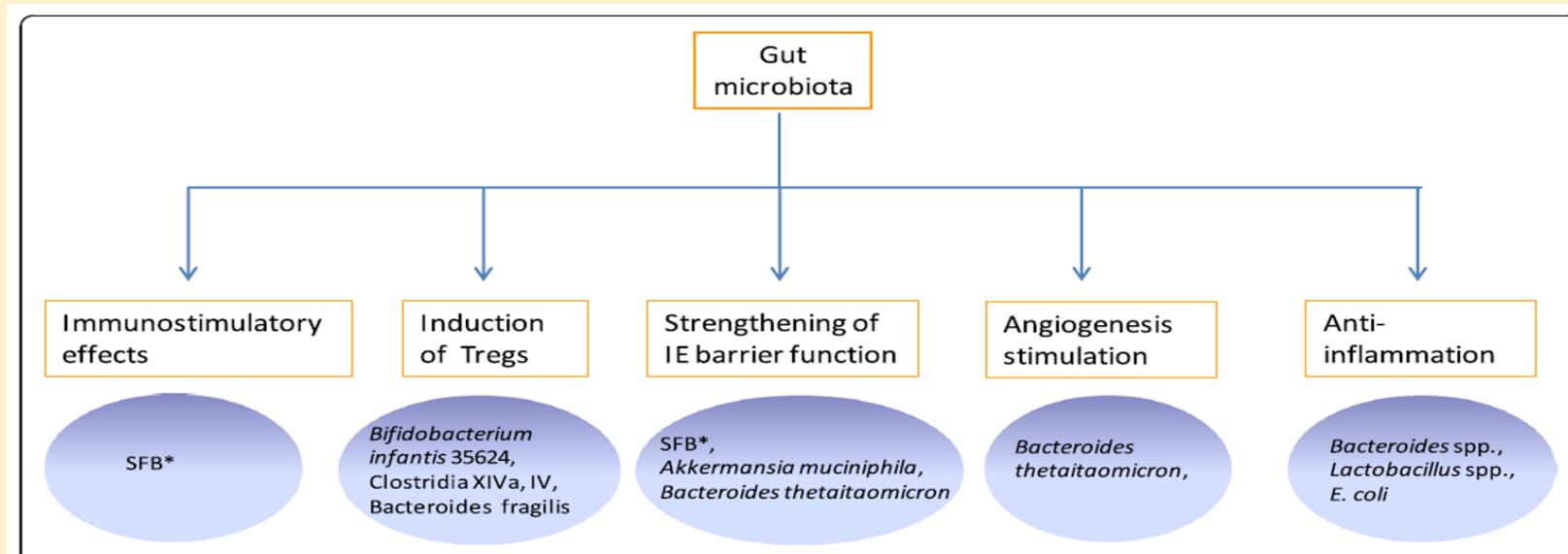
- ***Salmonella* genes that control oxidative metabolism are also linked to inflammasome activation in poultry**
- ***Salmonella* activation of NLRP3 inflammasome induces the development of a more effective protective immunity against a homologous *Salmonella* challenge**
- **Kinome peptide signal transduction pathway analysis:**
  - T cell receptor, B cell receptor, Insulin, HIF, and PI-3K signaling pathways
  - NLRP3 inflammasome activation = stronger immunometabolic protective mechanism(s)

## Future OSQR Plans (2)

- Evaluate cross protection against *Salmonella* serovars
- Evaluate vaccination of broiler breeders and effects of trans-generational stimulation of innate immunity
- Role for NLRP6 inflammasome = intestinal epithelium?

# Immunomodulatory commensal bacteria

Although numerous probiotic microorganisms have been identified, there remains a compelling need to discover organisms that elicit more robust therapeutic responses, are compatible with the host, and can affect a ***specific arm of the host immune system in a well-controlled, physiological manner***





# Spatial and Temporal Changes in the Broiler Chicken Cecal and Fecal Microbiomes and Correlations of Bacterial Taxa with Cytokine Gene Expression

Brian B. Oakley<sup>1\*</sup> and Michael H. Kogut<sup>2</sup>

<sup>1</sup> College of Veterinary Medicine, Western University of Health Sciences, Pomona, CA, USA, <sup>2</sup> United States Department of Agriculture, Agricultural Research Service, Southern Plains Area Research Center, College Station, TX, USA

To better understand the ecology of the poultry gastrointestinal (GI) microbiome and its interactions with the host, we compared GI bacterial communities by sample type (fecal or cecal), time (1, 3, and 6 weeks posthatch), and experimental pen (1, 2, 3, or 4), and measured cecal mRNA transcription of the cytokines IL18, IL1 $\beta$ , and IL6, IL10, and TGF- $\beta$ 4. The microbiome was characterized by sequencing of 16S rRNA gene

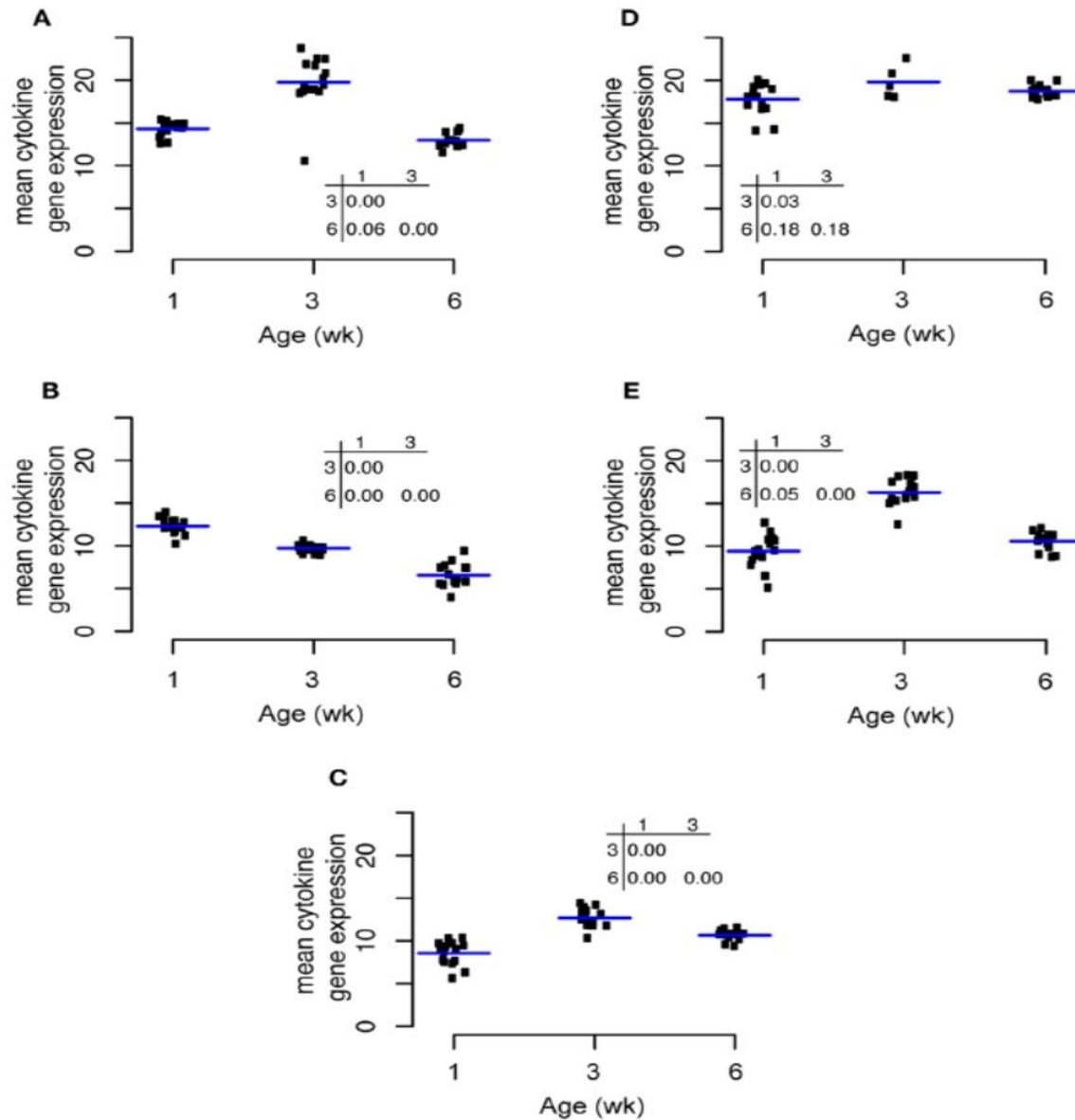


FIGURE 5 | Changes in cytokine expression through time for IL1 $\beta$  (A), IL6(B), IL18(C), TGF- $\beta$ 4 (D), and IL10 (E).

# Cecal Microbiome-Cytokine Correlations

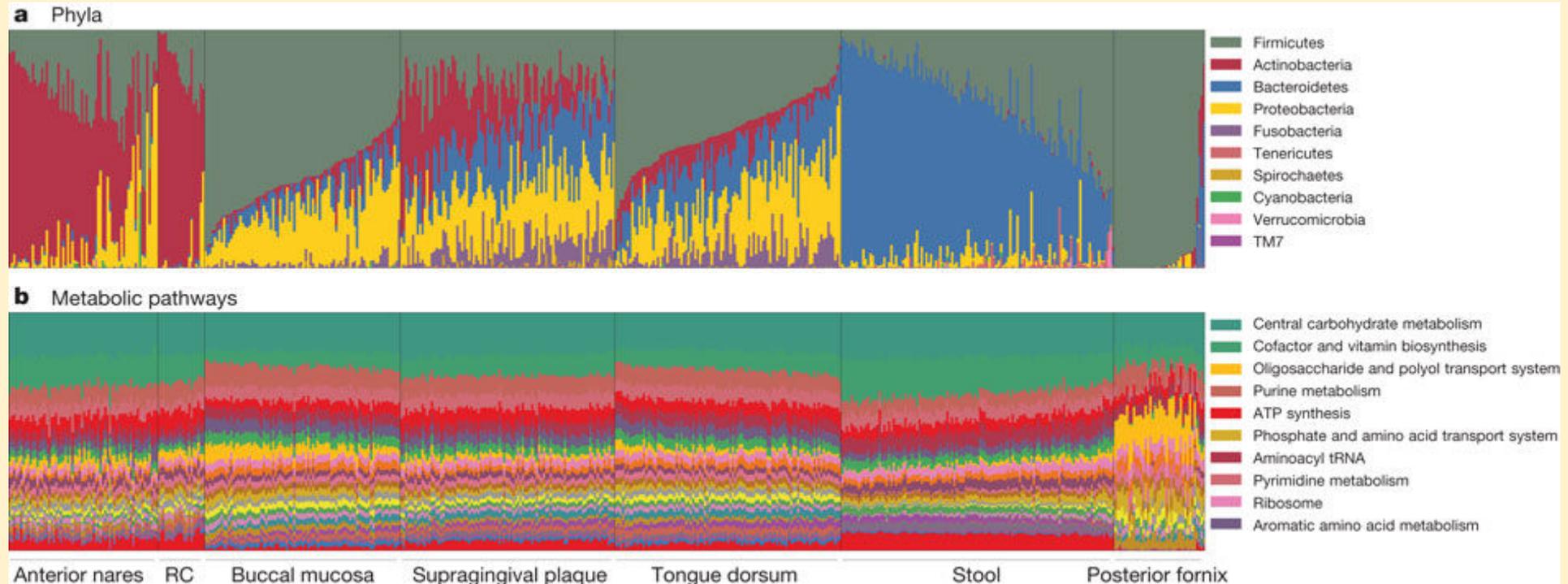
Transcription of pro-inflammatory cytokines was generally negatively correlated with the relative abundance of various members of the phylum Firmicutes and positively correlated with Proteobacteria.

1. **Proteobacteria** (phylum) were **positively correlated** with a pro-inflammatory response, most strongly with IL6 expression = *Escherichia/ Shigella, Parasutterella, Ruminococcus, and Vampirovibrio*.
2. **Firmicutes** were **negatively correlated** with pro-inflammatory (IL6) and positively correlated with anti-inflammatory (TGF- $\beta$ 4) cytokine expression, consistent with a growing body of evidence demonstrating positive influences of Firmicutes on gut health = *Faecalibacterium, Caloramator, Roseburia, and Clostridium*

# Impact (3)

1. Plasticity of microbiome and its involvement in host physiological pathways provides a desirable potential therapeutic target
2. **A better understanding of microbial ecology AND how interventions impact microbiota and host required**
3. **High through-put sequencing provides detailed survey of microbial community composition, but a **functional and mechanistic** understanding of the microbial ecology is required –identification of “functional groups”**
4. Consequences of manipulation of microbiome to improve poultry productivity (removing detrimental or enriching beneficial taxa) expected to go beyond improving feed conversion
  - ❖ **A disturbed microbiome may reveal commensals that have pathogenic potential**

# Future OSQR Plans (3)



**Carriage of microbial taxa varies over time**

**Metabolic pathways (function) remains stable**

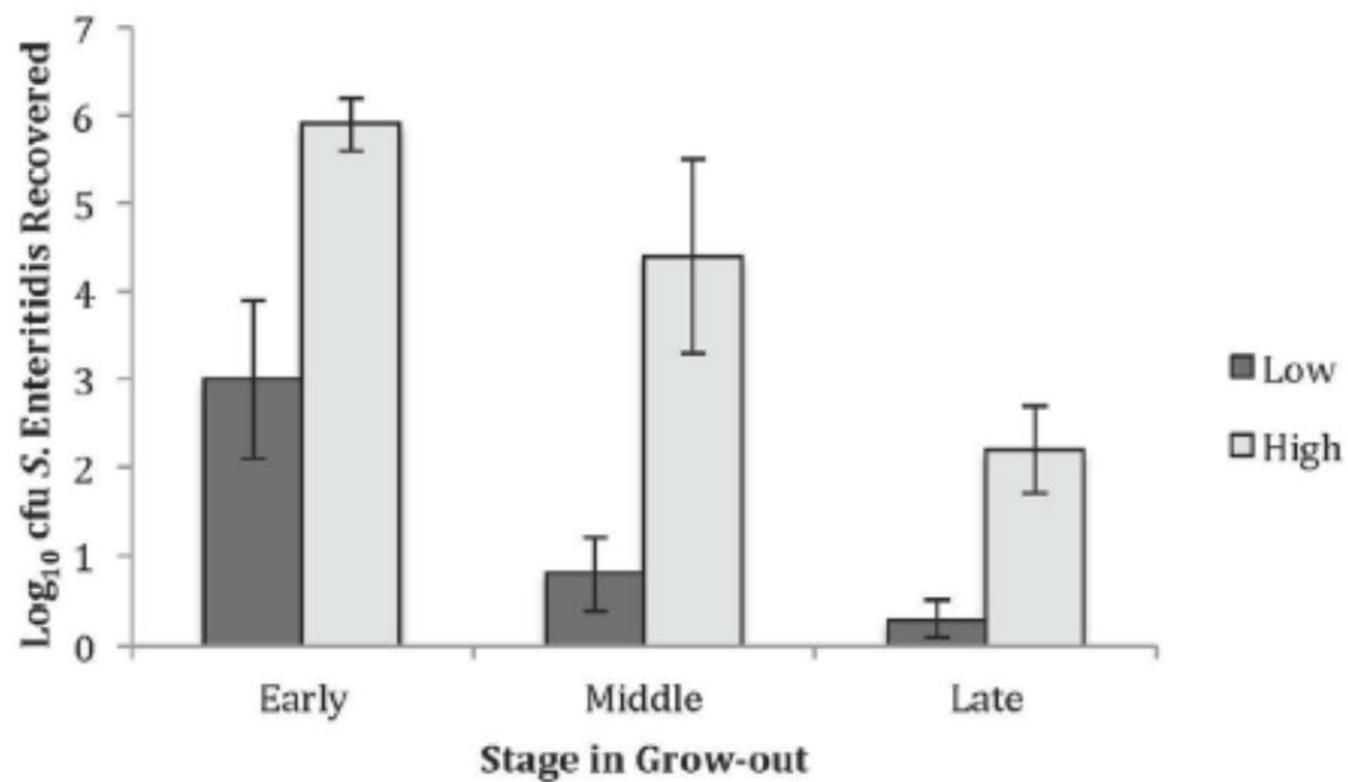


# Differential Levels of Cecal Colonization by *Salmonella* Enteritidis in Chickens Triggers Distinct Immune Kinome Profiles

Christina L. Swaggerty<sup>1\*</sup>, Michael H. Kogut<sup>1</sup>, Haiqi He<sup>1</sup>, Kenneth J. Genovese<sup>1</sup>, Casey Johnson<sup>2</sup> and Ryan J. Arsenault<sup>2</sup>

<sup>1</sup>U.S. Department of Agriculture, Agricultural Research Service, College Station, TX, United States, <sup>2</sup>Department of Animal and Food Sciences, University of Delaware, Newark, DE, United States

at each time were selected for kinomic analyses. Key biological processes associated with lower loads of *Salmonella* clustered around immune responses, including cell surface receptor signaling pathway, positive regulation of cellular processes, defense response, innate immune response, regulation of immune response, immune system process, and regulation of signaling. Further evaluation showed specific pathways including chemokine, Jak–Stat, mitogen activated protein kinase, and T cell receptor signaling pathways were also associated with increased resistance. Collectively, these findings demonstrate that it is possible to identify key mechanisms and pathways that are associated with increased resistance against *S. Enteritidis* cecal colonization in chickens. Therefore, providing a foundation for future studies to identify specific proteins within these pathways that are associated with resistance, which could provide breeders additional biomarkers to identify birds naturally more resistant to this important foodborne pathogen.



**FIGURE 1** | Recovered *Salmonella* Enteritidis in the high and low groups used for the peptide and antibody arrays. The average Log<sub>10</sub> colony forming units recovered for the low and high *S. Enteritidis* groups at the early, middle, and late times.

**TABLE 2** | Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways identified with the peptide array at the early stage infections in chickens with high and low levels of *Salmonella* Enteritidis colonization.

High <i>S. Enteritidis</i>	Number of peptides	Low <i>S. Enteritidis</i>	Number of peptides
B cell receptor signaling pathway	4	B cell receptor signaling pathway	4
Mitogen activated protein kinase (MAPK) signaling pathway	6	Chagas disease	4
		<b>Chemokine signaling pathway</b>	<b>4</b>
		Epithelial cell signaling pathway in <i>Helicobacter pylori</i> infection	2
		ErbB signaling pathway	6
		<b>Fc <math>\epsilon</math> RI signaling pathway</b>	<b>5</b>
		Fc- $\gamma$ receptor-mediated phagocytosis	5
		<b>Focal adhesion</b>	<b>6</b>
		GnRH signaling pathway	3
		<b>Insulin signaling pathway</b>	<b>7</b>
		<b>Jak-Stat signaling pathway</b>	<b>7</b>
		<b>MAPK signaling pathway</b>	<b>8</b>
		mTor signaling pathway	2
		Natural killer cell mediated cytotoxicity	6
		<b>Neurotrophin signaling pathway</b>	<b>6</b>
		Osteoclast differentiation	4
		<b>Pathways in cancer</b>	<b>10</b>
		<b>T cell receptor signaling pathway</b>	<b>6</b>
		Toll-like receptor signaling pathway	2
		Toxoplasmosis	3
		<b>Tuberculosis</b>	<b>3</b>
		VEGF signaling pathway	5

Pathways listed in bold showed statistically significant changes at all time points in birds with either low or high loads of *S. Enteritidis*.

**TABLE 3** | Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways identified with the peptide array at the middle stage infections in chickens with high and low levels of *Salmonella* Enteritidis colonization.

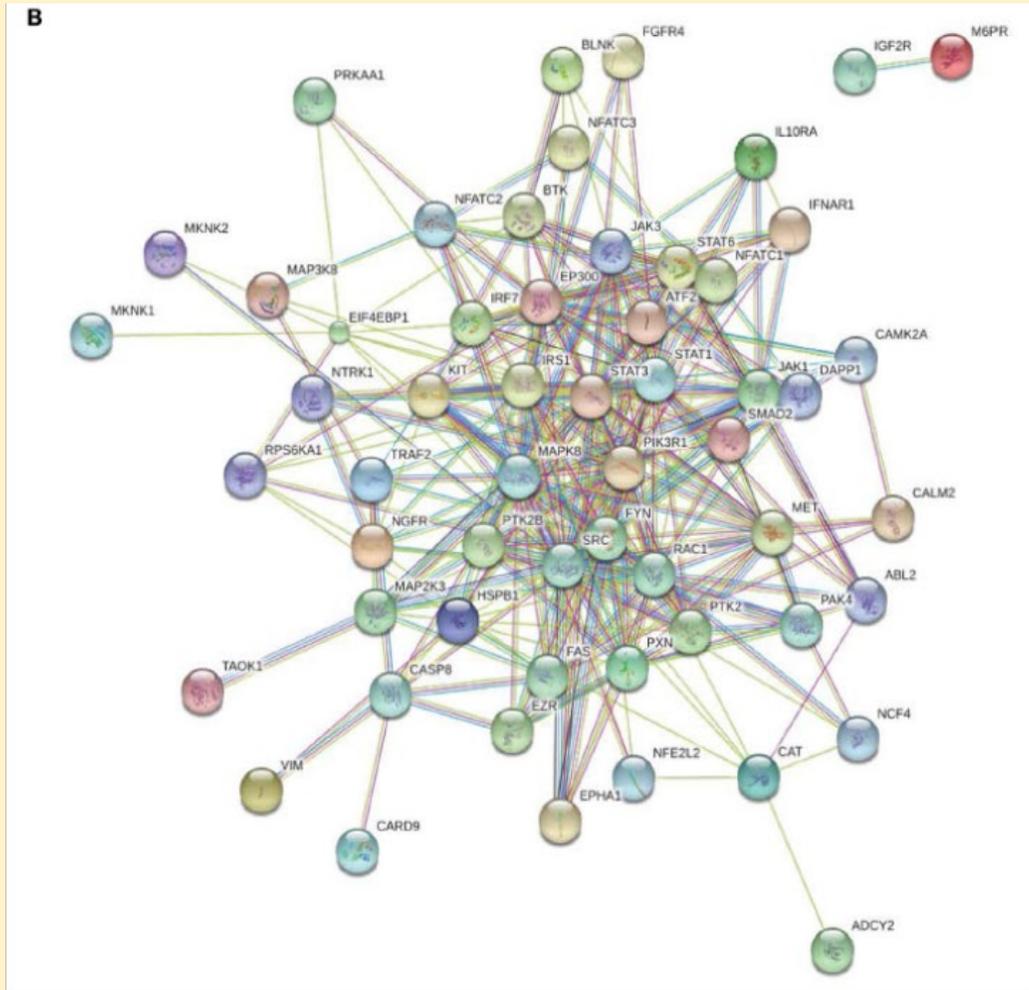
High <i>S. Enteritidis</i>	Number of peptides	Low <i>S. Enteritidis</i>	Number of peptides
Bacterial invasion of epithelial cells	5	<b>Chemokine signaling pathway</b>	<b>5</b>
Chemokine signaling pathway	7	ErbB signaling pathway	4
Chronic myeloid leukemia	4	<b>Fc <math>\epsilon</math> RI signaling pathway</b>	<b>2</b>
ErbB signaling pathway	7	<b>Focal adhesion</b>	<b>2</b>
Fc $\epsilon$ RI signaling pathway	1	GnRH signaling pathway	4
Focal adhesion	6	<b>Insulin signaling pathway</b>	<b>3</b>
Insulin signaling pathway	5	<b>Jak-Stat signaling pathway</b>	<b>4</b>
Mitogen activated protein kinase (MAPK) signaling pathway	6	Leukocyte transendothelial migration	2
Natural killer cell mediated cytotoxicity	4	<b>MAPK signaling pathway</b>	<b>4</b>
Neurotrophin signaling pathway	6	mTor signaling pathway	2
Pathways in cancer	6	<b>Neurotrophin signaling pathway</b>	<b>5</b>
T cell receptor signaling pathway	4	<b>Pathways in cancer</b>	<b>2</b>
		Regulation of actin cytoskeleton	1
		<b>T cell receptor signaling pathway</b>	<b>1</b>
		<b>Tuberculosis</b>	<b>3</b>

Pathways listed in bold showed statistically significant changes at all time points in birds with either low or high loads of *S. Enteritidis*.

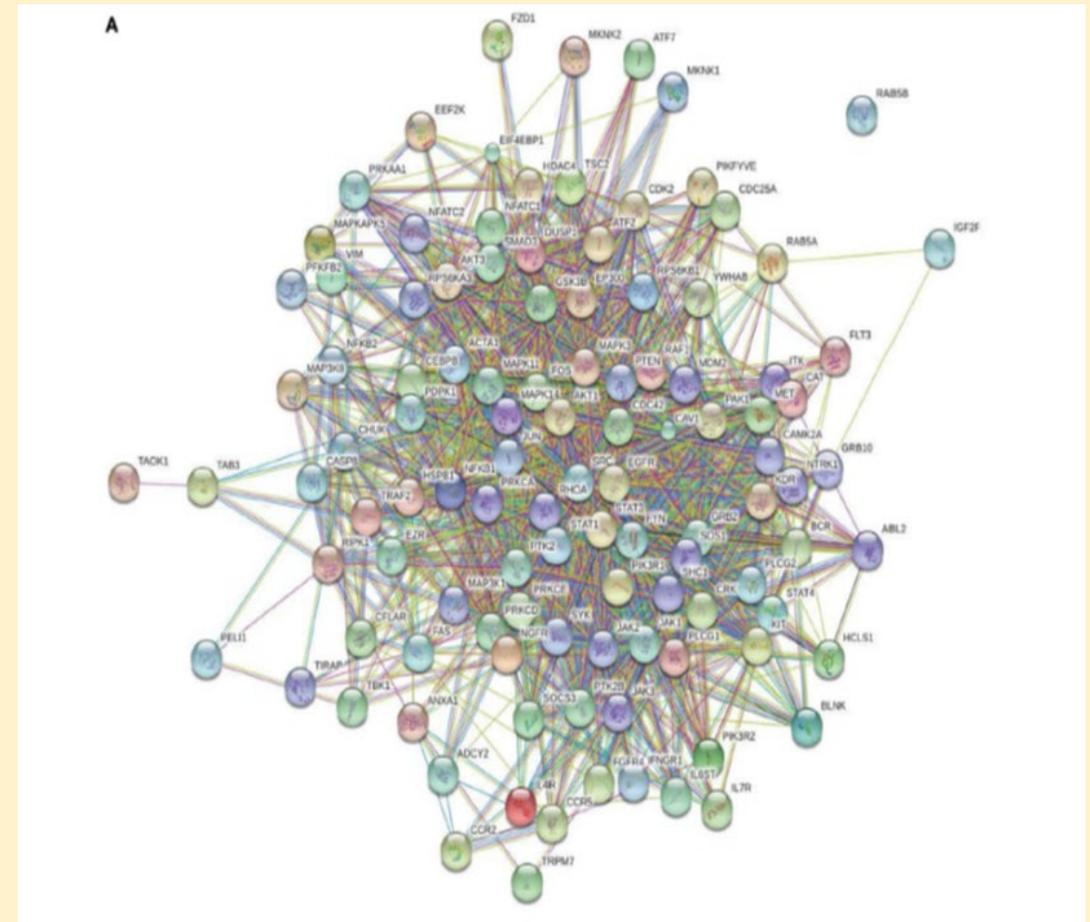
**TABLE 4** | Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways identified with the peptide array at the late stage infections in chickens with high and low levels of *Salmonella* Enteritidis colonization.

High <i>S. Enteritidis</i>	Number of peptides	Low <i>S. Enteritidis</i>	Number of peptides
Endocytosis	2	B cell receptor signaling pathway	3
ErbB signaling pathway	3	<b>Chemokine signaling pathway</b>	<b>5</b>
Focal adhesion	3	ErbB signaling pathway	3
Jak-Stat signaling pathway	2	<b>Fc <math>\epsilon</math> RI signaling pathway</b>	<b>3</b>
mitogen activated protein kinase (MAPK) signaling pathway	2	Fc- $\gamma$ R-mediated phagocytosis	2
Neurotrophin signaling pathway	2	<b>Focal adhesion</b>	<b>3</b>
Osteoclast differentiation	1	<b>Insulin signaling pathway</b>	<b>2</b>
Pathways in cancer	3	<b>Jak-Stat signaling pathway</b>	<b>3</b>
T cell receptor signaling pathway	1	<b>MAPK signaling pathway</b>	<b>1</b>
Toll-like receptor signaling pathway	2	Natural killer cell mediated cytotoxicity	2
Toxoplasmosis	2	<b>Neurotrophin signaling pathway</b>	<b>3</b>
Tuberculosis	3	Osteoclast differentiation	3
VEGF signaling pathway	2	<b>Pathways in cancer</b>	<b>5</b>
		<b>T cell receptor signaling pathway</b>	<b>2</b>
		Toll-like receptor signaling pathway	2
		Toxoplasmosis	2
		<b>Tuberculosis</b>	<b>2</b>
		VEGF signaling pathway	3

Pathways listed in bold showed statistically significant changes at all time points in birds with either low or high loads of *S. Enteritidis*.



Protein-protein interactions: early high SE load



Protein-protein interactions: early low SE load

# Impact (4)

- Identified individual birds within a population of a genetic line of birds that were either more resistant or susceptible to *Salmonella* colonization.
- Using a kinome array, identify key mechanisms and pathways associated with increased resistance against *Salmonella* colonization within population.
- Identified specific proteins associated with *Salmonella* colonization

# Future OSQR Plans (4)

- Dissect pathways associated with increased resistance to *Salmonella* colonization = identify individual proteins associated with the desired phenotype.
- Is there an interaction/interplay between the host's immune-metabolic pathways that contribute to increased resistance to *Salmonella* colonization
- Consider the role of the host microbiome in determining if a bird is more resistant or susceptible to *Salmonella* colonization.